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## Antifungal potentiality of leaves of some higher plants against *Rhizoctonia solani* causing Damping off disease of Brinjal

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A screening of leaves of 21 species of angiosperms was made for their volatile toxicity against *Rhizoctonia solani*. The volatile substances from *Callistemon lanceolatus* and *Citrus medica* were toxic against *R. solani*. Soil amendment with the leaves of *C. lanceolatus* and *C. medica* and the mixture of both the plants leaves controlled damping off disease of brinjal up to 61.11, 72.25 and 83.33 % respectively in soil infested with *R. solani*. Soil amendments with leaves and mixture of leaves of these fungitoxic plants increase the saprophytic fungal community .

**Key words** : Volatile toxicity, *Rhizoctonia solani*, soil amendment, damping off disease, fungicides, 21 angiospermic plants

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### INTRODUCTION

Synthetic chemicals with various degree of persistence are employed as fungicides in crop protection. The use of many such fungicides has now been cautioned due to their carcinogenicity, teratogenicity, and other residual toxicities (Bajaj and Ghosh, 1975; Sax, 1987; Arya, 1988; Lingk, 1991). Several of the synthetic fungicides are reported to cause adverse effect on treated soil ecosystem because of their non-biodegradable nature ( Shashikant *et al.*, 1989). Scientists are now looking for some alternatives for the control of plants diseases. In search of better alternatives natural products are considered to be environmentally safe for control of plant diseases (Beye, 1978). Higher plants in the tropics are reservoir of different secondary metabolites and provides an almost limitless source of useful chemicals with different biological properties (Sbragia, 1975). Several higher plants have been found to possess outstanding fungitoxicity against mycelial growth or spore germination of different phytopathogenic fungus *in vitro* (Pandey and Dubey, 1991, 1992, 1994; Lee, 2006). Therefore, in present piece of work the leaves extract of some plants have been screened for their volatile toxicity and soil amendment with

potent fungitoxic plants against *Rhizoctonia solani* causing damping off of brinjal have been evaluated.

### MATERIALS AND METHODS

Fresh leaves of 21 angiosperms were screened for their volatiles against the test fungus *R. solani*. The leaves of ( 20 g) of each plant were thoroughly washed with 70% ethanol and finally with sterile distilled water in pestle and mortar (1:1, w/v) and filtered through double layered sterilized cheese cloth. The clear extract (aqueous extract) thus obtained was assayed for its antifungal activity by the inverted Petriplate Technique (Peach and Tracy, 1955). Potato dextrose agar medium was aseptically poured in Petriplates (10 ml / plate) and was inoculated with discs cut from 7 days old culture of the test fungus. The inoculated plates were inverted upside down. Then 5 ml of the prepared extract was aseptically pipetted to the lid of the Petri plate. Control sets were prepared similarly using 5 ml sterilized distilled water. The plates were kept at 25±2°C for 7 days. The experiment was run triplicate. The fungitoxicity was calculated as per formula : %mycelial growth inhibition = 100 (dc- dt)/dc where dc = mean colony diameter of control; dt = mean colony diameter of treatment



Amendment of soil with leaves of *C. lanceolatus* and *C. medica* was done (Pandey and Dubey, 1997) to find out the potentiality of leaves in control of damping-off disease of *Solanum melongena* caused by *R.solani*. Thirty kg of garden soil was collected for setting treatments and controls. Six kg of the soil was filled in 3 earthen pots (2 kg/pot) which served as uninoculated control. Twenty four kg of the soil was inoculated with 120 g inoculum of *R.solani* maintained on oat sand. The soil inoculated with *R.solani* was filled (2 kg/pot) in to 12 pots separately and kept for one week to establish the infection. Three of infested pot served as inoculated controls. The remaining pots, were amended with leaves pieces (0.25cm<sup>2</sup>) of *C. lanceolatus*, *C. medica* and mixture of leaves pieces (1:1 w/w) of these plants @ 20 g/pot ( 3 pots for each) separately. Twenty days after the amendment, the seeds of brinjal soaked for 6 h in sterilized water, were sown (2 cm deep ) equidistantly in all 15 pots @ 20 seeds/pot. Experiments were repeated thrice. After 14 days of sowing at two leaf stage the per cent seedling mortality and per cent disease control were calculated by the following formulae (Kataria and Grover, 1976) : % seedling mortality = 100 – Seedling stand in inoculated soil/ Seedling stand in uninoculated soil × 100% disease control = 100 – % disease in treatment set/% disease in treatment control × 100

The effects of soil amendment with leaves of *C. lanceolatus*, *C. medica* and mixture of leaves of these plants on soil mycoflora were investigated by the method of Khalis and Manoharachari (1985 ).Sterilized earthen pots were filled with 1 kg of garden soil. The soil was amended with 5, 10, and 15% of leaf pieces of young leaves ( 0.5 cm<sup>2</sup> ) of *C. lanceolatus*, *C.medica* and mixture of both the plant pieces (1:1 w/v) separately. Controls sets contained unamended soil. The pots were watered regularly with equal amount of sterilized water. After 30 days, all the sets were subjected to mycoflora analysis separately by the Waksman's dilution plate method (Waksman, 1952) as well as Warcup's soil plate method (Dhingra and Sinclair, 1986) using Martin agar medium. The soil from the top 1 cm was removed in a glass container. The container was kept in a water bath for 30 minutes at 60°C and mycoflora analysis was made.

## RESULTS AND DISCUSSION

During screening of leaves of angiospermic plants,

most plant species showed either poor ( below 50%) or moderate ( above 50% and below 100% ) fungitoxicity. However, leaf extracts of *C. lanceolatus* and *C.medica* inhibited the growth of test fungus completely (Table 1). In pot experiments soil amended with leaves of *C. lanceolatus*, *C.medica* and mixture of both the plant leaves showed control of damping – off by 61.11, 72.25 and 83.33% respectively in soil infested with *R. solani* (Table 2) .The number of fungal types greatly increased as a result of soil amendment with the leaves and the mixture of leaves of the test plants.The soil amended with 5, 10, and 15% of *Callistemone* leaves harboured 12, 11 and 11 saprophytic species respectively. By amendment with 5,10 and 15% *Citrus* leaves 10, 9 and 9 species of fungi were isolated respectively. While the mixture of both the plant leaves harboured 10 saprophytic species in each set. Increase the percentage occurrence of fungi such as *Trichoderma harzianum* and *T. viride* observed in amended soils (Table 3). In unamended set 24 fungal species were isolated. Moreover in amended soils the fungus namely *Botrytis cineria* completely disappeared. Some fungi namely *Alternaria*

**Table 1** : Fungitoxicity of volatile compounds of leaves of some higher plants against *Rhizoctonia solani* (±S.E. of the Mean)

Plant tested	% inhibition of growth <i>Rhizoctonia solani</i>
<i>Acacia arabica</i>	45.2±0.04
<i>Albizia labbeck</i>	46.3±0.03
<i>Annona squamosa</i>	43.4±0.05
<i>Antirrhinum orontium</i>	42.3±0.06
<i>Artocarpus heterophyllus</i>	23.3±0.03
<i>Callistemone lanceolatus</i>	100
<i>Cannabis sativus</i>	75.5±0.04
<i>Cassia nodosa</i>	35.5±0.03
<i>Citrus medica</i>	100
<i>Daicas carpta</i>	24.3±0.06
<i>Gomphrena globosa</i>	19.5±0.07
<i>Ipomea fistulosa</i>	32.2±0.05
<i>Morus indica</i>	54.3±0.04
<i>Phyllanthus emblica</i>	65.2±0.02
<i>Polyalthia longifolia</i>	44.1±0.03
<i>Psidium guajava</i>	43.3±0.04
<i>Punica granatum</i>	63.3±0.05
<i>Raphanus sativus</i>	63.6±0.06
<i>Ranunculus scleratus</i>	73.8±0.07
<i>Saraca indica</i>	26.2±0.07
<i>Ziziphus mauritiana</i>	33.3±0.06



**Table 2 :** Soil amendment with leaves of *Callistemone lanceolatus*, *Citrus medica* and mixture of leaves of these plants (1:1 w/w) for control of damping-off disease of brinjal (*Solanum melongena*) caused by *Rhizoctonia solani*.

Pathogen	Average no. of healthy seedlings					% seeding mortality			% disease control			
	control		Treatment		Control	Treatment	C.L.		C.M.	C.L.±C.M.		
	Unino.	Ino.	C.L.	C.M.	C.L±C.M.	C.L.	C.M.	C.L±C.M.				
<i>Rhizoctonia solani</i>	25±0.03	07±0.02	18±0.01	20±0.01	22±0.04	72±0.03	28±0.01	20±0.01	12±0.01	61.11±0.03	72.25±0.02	82.33±0.01

**Table 3 :** Per cent occurrence of fungi in soil amended with leaves of *Callistemone lanceolatus* and *Citrus medica* and combination of both the plant leaves

Fungi	% Occurrence of fungi									
	Leaves amended (w/w)									
	<i>C.lanceolatus</i>				<i>C.medica</i>			<i>C. lanceolatus±C.medica</i>		
	Control	5%	10%	15%	5%	10%	15%	5%	10%	15%
<b>Fungi stimulated</b>										
<i>Aspergillus flavus</i>	14±0.05	14±0.02	20±0.02	25±0.01	15±0.03	22±0.01	30±0.03	16±0.01	25±0.03	35±0.04
<i>A.fumigatus</i>	07±0.02	06±0.01	08±0.03	10±0.02	07±0.02	10±0.03	12±0.01	09±0.01	12±0.01	15±0.01
<i>A. luchuensis</i>	04±0.03	06±0.02	06±0.03	08±0.04	08±0.01	10±0.01	15±0.01	08±0.03	14±0.01	16±0.02
<i>A. parasiticus</i>	03±0.02	03±0.01	04±0.02	06±0.03	ND	ND	ND	06±0.03	08±0.01	12±0.01
<i>Botrytis cineria</i>	02±0.03	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>Penicillium citrinum</i>	10±0.05	12±0.01	14±0.01	16±0.02	13±0.01	16±0.01	18±0.01	14±0.01	16±0.03	18±0.03
<i>P. digitatum</i>	09±0.02	10±0.02	12±0.01	14±0.01	12±0.03	13±0.02	15±0.01	16±0.01	18±0.01	20±0.01
<i>P. italicum</i>	07±0.01	09±0.01	10±0.03	11±0.02	09±0.03	12±0.01	14±0.02	12±0.01	18±0.03	20±0.01
<i>P. vermiculatum</i>	04±0.03	04±0.01	05±0.02	05±0.01	06±0.01	08±0.01	10±0.02	07±0.01	10±0.02	15±0.01
<i>Trichoderma viride</i>	09±0.01	11±0.03	14±0.03	18±0.01	13±0.01	16±0.01	20±0.01	15±0.01	20±0.03	25±0.01
<i>T. harzianum</i>	07±0.02	09±0.01	11±0.02	15±0.01	10±0.02	11±0.02	15±0.02	12±0.02	16±0.02	20±0.03
<b>Fungi affected</b>										
<i>Aspergillus niger</i>	10±0.03	08±0.01	06±0.02	02±0.01	03±0.01	ND	ND	ND	ND	ND
<i>A. sulphureus</i>	03±0.02	01±0.01	ND	ND	ND	ND	ND	ND	ND	ND
<i>Alternaria brassicae</i>	04±0.03	03±0.01	02±0.01	ND	02±0.03	01±0.01	ND	ND	ND	ND
<i>A. raphani</i>	04±0.02	04±0.02	03±0.02	ND	03±0.01	01±0.01	ND	ND	ND	ND
<i>Cercospora cajani</i>	05±0.01	03±0.01	02±0.03	02±0.01	03±0.02	02±0.02	ND	ND	ND	ND
<i>C. capsici</i>	07±0.02	05±0.03	04±0.02	02±0.02	04±0.02	02±0.03	ND	ND	ND	ND
<i>Cladosporium capsici</i>	10±0.03	09±0.01	08±0.02	03±0.01	07±0.01	05±0.01	03±0.01	08±0.01	05±0.02	02±0.03
<i>Colletotricum capsici</i>	08±0.02	08±0.01	07±0.02	05±0.01	07±0.01	05±0.01	03±0.01	04±0.01	03±0.01	01±0.02
<i>Drechslera graminea</i>	07±0.02	06±0.02	03±0.01	01±0.01	06±0.02	02±0.01	ND	ND	ND	ND
<i>Fusarium semitectum</i>	06±0.02	04±0.01	02±0.01	ND	ND	ND	ND	ND	ND	ND
<i>Pythium debaryanum</i>	07±0.01	07±0.01	05±0.02	03±0.01	05±0.01	03±0.01	ND	ND	ND	ND
<i>P. proliferum</i>	05±0.07	03±0.02	ND	ND	ND	ND	ND	ND	ND	ND
<i>Rhizoctonia solani</i>	06±0.01	03±0.01	02±0.02	01±0.02	03±0.02	ND	ND	ND	ND	ND

ND = Not detected.

*brassicae*, *A. raphani*, *Cercospora cajani*, *C.capsici*, *Drechslera graminea*, *Fusarium semitectum*, *Pythium debaryanum*, *P. proliferum* and *Rhizoctonia solani* disappeared in leaves mixture of both the plants. These findings indicate that for the management of disease caused by *R.solani* the leaves of *C. lanceolatus* and *C.medica* may be used. The increase in number of some saprophytic fungi in soil amended with leaves and leaves mixture of the test

plants may be an additional merit in soil disease control. These saprophytic fungi may provide disease control through antagonism.

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## REFERENCES

- Arya, A. 1988. Control of phomopsis fruit rot by leaf extracts of certain medicinal plants (including microbes and fungi). In *Indigenous Medicinal Plants* (Purshotam Kausik Ed.) Today and Tomorrow's Printers and Publishers, New Delhi. pp. 41-46.
- Bajaj, B. S. and Ghosh, A. K. 1975. Antifungal antibiotics in perspective. In *Advances in Mycology and Plant Pathology* (S. P. Raychoudhuri, A. Verma, K.S. Bhargava and B. S. Mehrotra, Eds) Sagar Printers, New Delhi, pp. 297-309.
- Beye, F. 1978. Insecticides from vegetable kingdom. *Plant Res. Develop.* **7** : 13-31.
- Dhingra, O. D. and Sinclair, J. B. 1986. *Basic Plant Pathology Methods*. CRC Press London.
- Kataria, H. R. and Grover, R. K. 1976. Some factors affecting the control of *Rhizoctonia solani* by systemic and non-systemic fungicides. *Annals of Applied Biology.* **82** : 267-278.
- Khalish, N. and Manoharachari, C. 1985. Studies on mycoflora changes in soil cake amended and unamended soils. *Indian Phytopathology*, **38** : 462-466.
- Lee, H. S. 2006. Fungicidal properties of active components from *Acorus gramineus rhizome* against phytopathogenic fungi. *Bio Resource Technology*. (In press)
- Lignk, W. 1991. Health risk evaluation of pesticide contaminations in drinking water. *Gesunde Pflangen.* **43** : 21-25.
- Paech, K. and Tracey, M. V. 1955. *Modern Methods of Plant Analysis*. Vol. III. Springer-Verlag, Berlin.
- Pandey, V. N. and Dubey, N. K. 1991. The synergistic activity of volatile fungitoxic compound from some higher plants. *Acta Botanica Indica.* **19** : 290-295.
- Pandey, V. N. and Dubey, N. K. 1992. Effect of essential oils from some higher plants against fungi causing damping-off disease. *Biologia Plantarum.* **34**(1-2): 143-147.
- Pandey, V. N. and Dubey, N. K. 1994. Antifungal potential of leaves and essential oils from higher plants against soil phytophogens. *Soil. Biol. Biochem.* **26**(10): 1417-1421.
- Pandey, V. N. and Dubey, N. K. 1997. Antifungal potentiality of some higher plants against *Pythium* species causing damping-off in tomato. *Nat. Acad. Sci. Letters.* **20**(5&6): 68-70.
- Sax, N. I. 1987. Hazardous chemicals Information Annual No. 2. Reinhold Company, New York.
- Sbragia, R. J. 1975. Chemical control of plant diseases, an exciting future. *Annual Review of Phytopathology.* **13** : 257-267.
- Shashikant, Vohra, S. and Sahai, Y. N. 1989. *Trends in Environmental Pollution and Pesticide Toxicology*. Jagmander Book Agency, New Delhi.
- Waksman, A. S. 1952. *Soil Microbiology*. John Wiley and Sons, New York.